

**Thiamethoxam**

**Thiamethoxam SL (A23943A) - *Salmonella Typhimurium* and  
*Escherichia Coli* Reverse Mutation Assay**

**Final Report**

**TEST GUIDELINE(S):** OECD 471 (2020)

**AUTHOR(S):** Dr. Steffi Chang

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**PERFORMING LABORATORY:** ICCR-Roßdorf GmbH  
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Bracknell, Berkshire RG42 6EY, United Kingdom

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This study performed in the test facility of ICCR-Rosßdorf GmbH, In den Leppsteinswiesen 19, 64380 Rosßdorf, Germany was conducted in compliance with Good Laboratory Practice Regulations:

Chemikaliengesetz (Chemicals Act) of the Federal Republic of Germany, "Anhang 1" (Annex 1), in its currently valid version

OECD Principles of Good Laboratory Practice, (as revised in 1997), ENV/MC/CHEM(98)17

EC Commission Directive 2004/10/EC

These procedures are compatible with Good Laboratory Practice regulations specified by regulatory authorities throughout the European Community, the United States (EPA and FDA), and Japan (MHW, MAFF, and METI), and other countries that are signatories to the OECD Mutual Acceptance of Data Agreement.

There were no circumstances that may have affected the quality or integrity of the study.

Dr. Steffi Chang  
Study Director Bacterial Systems

  
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Date: 14 June 2022

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## **FLAGGING STATEMENT**

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## QUALITY ASSURANCE STATEMENT

ICCR Study Number: 3977311  
Test substance: Thiamethoxam SL (A23943A)  
Study director: Dr. Steffi Chang  
Study Title: Thiamethoxam SL (A23943A) -  
*Salmonella Typhimurium* and  
*Escherichia Coli* Reverse Mutation Assay

Study based activities at the Test Facility ICCR-Roßdorf GmbH were audited and inspected. The details of these audits and inspections are given below.

Type of Inspection	Date(s) of Inspection	Date Reporting to Study Director, Test Facility Management
Study Plan Verification	16 December 2021	16 December 2021
Process – based Assessment of Response	13 January 2022	13 January 2022
Report Audit	22 March 2022	24 March 2022

General facilities and activities where this study was conducted were inspected on an annual basis and results are reported to the relevant responsible person and Management.

The statement is to confirm, that this report reflects the raw data.

  
\_\_\_\_\_  
Sabine Ebert

Quality Assurance Auditor  
ICCR-Roßdorf GmbH

  
\_\_\_\_\_  
Date

## PROJECT STAFF SIGNATURE

Study Director

Dr. Steffi Chang



.....  
Date: 14 June 2022



## GENERAL INFORMATION

### Contributors

The following contributed to this report in the capacities indicated:

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### Study Dates

Study initiation date:	17 December 2021
Experimental start date:	04 January 2022
Experimental completion date:	24 January 2022

### Deviations from the Guidelines

None

### Retention of Samples

None

### Performing Laboratory Test Substance Reference Number

S 2205211

### Other

ICCR-Roßdorf GmbH will archive:

Records and documentation relating to this study will be maintained in the archives of ICCR-Roßdorf GmbH for a period of 4 years from the date on which the Study Director signs the final report. This will include electronic and paper raw data, and report that support the reconstruction of the study.

At termination of the aforementioned period, the records and documentation will be transferred to the GLP compliant Archive of Rhenus Archiv Services GmbH, Frankfurt am Main for further archiving up to a total archiving period of 15 years.

A sample of the test substance will not be archived.

ICCR Roßdorf GmbH will retain in its archive a copy of the study plan and final report, and any amendments indefinitely.

### Deviations from the study plan

There were no deviations (unplanned changes) from the study plan.

**Distribution of the report**

Sponsor	2 electronic copies (1 pdf-file, 1 word-file)
Study Director	1 (original)



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## **1.0 EXECUTIVE SUMMARY**

### **1.1 Study Design**

This study was performed to investigate the potential of thiamethoxam SL (A23943A) to induce gene mutations in the plate incorporation test (Experiment I) and the pre-incubation test (Experiment II) using the *Salmonella typhimurium* (*S. typhimurium*) strains TA1535, TA1537, TA98, and TA100, and the *Escherichia coli* (*E. coli*) strains WP2 *uvrA* (pKM101) and WP2 (pKM101).

### **1.2 Results**

The plates incubated with the test substance showed normal background growth up to the maximal concentration of 5000 µg/plate with and without S9 mix in all strains used.

No cytotoxic effects, evident as a reduction in the number of revertants (below the indication factor of 0.5), occurred in all strains with and without metabolic activation.

No relevant increase in revertant colony numbers of any of the six tester strains was observed following treatment with thiamethoxam SL (A23943A) at any concentration, neither in the presence nor absence of metabolic activation (S9 mix). There was also no observed tendency of higher mutation rates with increasing concentrations in the range below the generally acknowledged border of biological relevance.

Appropriate reference mutagens were used as positive controls, which showed a distinct increase of induced revertant colonies consistent with the laboratory's historical control data demonstrated the sensitivity of the test system and the efficacy of the S9 mix. Each batch of S9 was also tested with 2 pro-mutagens, benzo(a)pyrene and 2-aminoanthracene.

### **1.3 Conclusion**

In conclusion, it can be stated that during the described mutagenicity tests and under the experimental conditions reported, thiamethoxam SL (A23943A) did not induce gene mutations by base pair changes or frameshifts in the genome of the strains used.

Therefore, thiamethoxam SL (A23943A) is considered to be negative (i.e. non-mutagenic) in the *Salmonella typhimurium* and *Escherichia coli* reverse mutation assay.

## 2.0 INTRODUCTION

### 2.1 Purpose

These experiments were performed to assess the potential of the test substance to induce gene mutations by means of the *S. typhimurium* and *E. coli* reverse mutation assay. Experiment I was performed as a plate incorporation assay. Since a negative result was obtained in this experiment, Experiment II was performed as a pre-incubation assay.

The most widely used assays for detecting gene mutations are those using bacteria (1). They are relatively simple and rapid to perform, and give reliable data on the ability of an agent to interact with DNA and produce mutations.

Reverse mutation assays determine the frequency with which an agent reverses or suppresses the effect of the forward mutation. The genetic target presented to an agent is therefore small, specific and selective. Several bacterial strains, or a single strain with multiple markers are necessary to assure reliable detection of mutagens that may be specific to one tester strain or locus. The reversion of bacteria from growth-dependence on a particular amino acid to growth in the absence of that amino acid (reversion from auxotrophy to prototrophy) is the most widely used marker.

The *S. typhimurium* histidine (his) and the *E. coli* tryptophan (trp) reversion system measures  $\text{his}^- \rightarrow \text{his}^+$  and  $\text{trp}^- \rightarrow \text{trp}^+$  reversions, respectively. The *S. typhimurium* and *E. coli* strains are constructed to differentiate between base pair (TA1535, TA100, WP2 *uvrA* (pKM101), and WP2 (pKM101)) and frameshift (TA1537, TA98) mutations.

According to the direct plate incorporation and pre-incubation method the bacteria are exposed to the test substance with and without metabolic activation and plated on selective medium. After a suitable period of incubation, revertant colonies are counted.

To establish a concentration response effect at least six concentrations with adequately spaced intervals were tested. The maximum concentration was 5000 µg/plate.

To validate the test, reference mutagens were tested in parallel to the test substance.

### 2.2 Test Guideline(s)

This study followed the procedures indicated by the following internationally accepted guideline and recommendations:

“Ninth Addendum to OECD Guidelines for Testing of Chemicals”, Section 4, No. 471:  
“Bacterial Reverse Mutation Test”, corrected June 26, 2020

### 3.0 MATERIALS AND METHODS

#### 3.1 Test Substance

Information as provided by the Sponsor.

Identification:	Thiamethoxam SL (A23943A)
Batch:	NSI001-085-017
Content of Thiamethoxam:	6.63% w/w corresponding to 75.42 g/L
Appearance:	Light yellow*, liquid
Recertification Date:	10 December 2023
Storage Conditions:	At room temperature
Stability in Solvent:	Not indicated by the Sponsor

\* determined by ICCR Roßdorf GmbH staff

The test substance concentrations were not adjusted for the content of Thiamethoxam SL (A23943A).

On the day of the experiment (immediately before use), the test substance was dissolved in deionised water. The solvent was chosen as the most suitable solvent compared to DMSO and ethanol, according to its solubilisation properties and its relative non-toxicity to the bacteria (2).

All formulations were prepared freshly before treatment and used within two hours of preparation. The formulation was assumed to be stable for this period unless specified otherwise by the Sponsor.

## 3.2 Controls

### 3.2.1 Negative controls

Concurrent untreated and solvent controls were performed.

### 3.2.2 Positive control substances

#### Without metabolic activation

Strains:	TA1535, TA100
Name:	Sodium azide, (NaN <sub>3</sub> )
Supplier:	Sigma-Aldrich, 82024 Taufkirchen, Germany
Batch No.:	STBJ7813
Purity:	≥ 99%
Dissolved in:	Deionised water
Concentration:	10 µg/plate
Strains:	TA1537, TA98
Name:	4-nitro-o-phenylene-diamine, (4-NOPD)
Supplier:	Sigma-Aldrich, 82024 Taufkirchen, Germany
Batch No.:	MKBM 5257V
Purity:	≥ 98%
Dissolved in:	DMSO (purity >99 %, Fisher Leics LE11 5RG, United Kingdom)
Concentration:	10 µg/plate in strain TA 98, 50 µg/plate in strain TA 1537
Strains:	WP2 <i>uvrA</i> (pKM101), WP2 (pKM101)
Name:	Methyl methane sulfonate, (MMS)
Supplier:	Sigma-Aldrich, 82024 Taufkirchen, Germany
Batch No.:	MKCL 6261
Purity:	≥ 99%
Dissolved in:	Deionised water
Concentration:	2.0 µL/plate

#### With metabolic activation

Strains:	TA1535, TA1537, TA98, TA100, WP2 <i>uvrA</i> (pKM101), WP2 (pKM 101)
Name:	2-aminoanthracene, (2-AA)
Supplier:	Sigma-Aldrich, 82024 Taufkirchen, Germany
Batch No.:	STBG 0630V
Purity:	≥ 96%
Dissolved in:	DMSO (purity > 99 %, Fisher Leics LE11 5RG, United Kingdom)
Concentration:	2.5 µg/plate (TA1535, TA1537, TA98, TA100), 10 µg/plate (WP2 <i>uvrA</i> (pKM101), WP2 (pKM101))

The stability of the positive control substances in solution is unknown but a mutagenic response in the expected range is sufficient evidence of biological activity.

### 3.3 Experimental Design

#### 3.3.1 Characterisation of the *Salmonella typhimurium* and *E. coli* strains

The histidine dependent strains are derived from *S. typhimurium* strain LT2 through mutations in the histidine locus. Additionally, due to the "deep rough" (*rfa*<sup>-</sup>) mutation they possess a faulty lipopolysaccharide envelope which enables substances to penetrate the cell wall more easily. A further mutation causes a reduction in the activity of an excision repair system. The last alteration includes mutational processes in the nitrate reductase and biotin genes produced in a UV-sensitive area of the gene named *uvrB*<sup>-</sup>. In the strains TA98 and TA100 the R-factor plasmid pKM101 carries the ampicillin resistance marker (3).

Strain WP2 (4) and its derivatives all carry the same defect in one of the genes for tryptophan biosynthesis. Tryptophan-independent (*Trp*<sup>+</sup>) mutants (revertants) can arise either by a base change at the site of the original alteration or by a base change elsewhere in the chromosome so that the original defect is suppressed. This second possibility can occur in several different ways so that the system seems capable of detecting all types of mutagen which substitute one base for another. Additionally, the *uvrA* derivative is deficient in the DNA repair process (excisable repair damage). Such a repair-deficient strain may be more readily mutated by agents. The *E. coli* strains WP2 *uvrA* (pKM101) and WP2 (pKM101) are constructed by introduction of the R-factor plasmid pKM101.

When summarized, the mutations of the *S. typhimurium* and *E. coli* strains used in this study can be described as follows:

Strains	Genotype	Type of mutations indicated
<i>Salmonella typhimurium</i>		
TA1537	<i>his</i> C 3076; <i>rfa</i> <sup>-</sup> ; <i>uvrB</i> <sup>-</sup>	frame shift mutations
TA98	<i>his</i> D 3052; <i>rfa</i> <sup>-</sup> ; <i>uvrB</i> <sup>-</sup> ; R-factor	" "
TA1535	<i>his</i> G 46; <i>rfa</i> <sup>-</sup> ; <i>uvrB</i> <sup>-</sup>	base-pair substitutions
TA100	<i>his</i> G 46; <i>rfa</i> <sup>-</sup> ; <i>uvrB</i> <sup>-</sup> ; R-factor	" "
<i>Escherichia coli</i>		
WP2 <i>uvrA</i> (pKM101)	<i>trp</i> E 56 <i>uvrA</i> <sup>-</sup> ; R-factor	base-pair substitutions and others
WP2 (pKM101)	<i>trp</i> E 56; R-factor	" "

Regular checking of the properties of the *S. typhimurium* and *E. coli* strains regarding the membrane permeability and ampicillin resistance; UV sensitivity, and amino acid requirement as well as normal spontaneous mutation rates is performed by ICCR-Roßdorf GmbH according to Ames *et al.* (5), Maron and Ames (3), and Mortelmans and Riccio (7). In this way it is ensured that the experimental conditions set down by Ames are fulfilled.

The bacterial strains TA1535, TA1537, TA98, TA100, WP2 *uvrA* (pKM101), and WP2 (pKM101) were obtained from Trinova Biochem GmbH (35394 Gießen, Germany).

### 3.3.2 Storage

The strain cultures were stored as stock cultures in ampoules with nutrient broth + 5 % DMSO (Fisher Leics, LE11 5RG, United Kingdom) in liquid nitrogen.

### 3.3.3 Precultures

The thawed bacterial suspension was transferred into 250 mL Erlenmeyer flasks containing nutrient medium (50 mL). A solution of ampicillin (50 µL, 25 µg/mL) was added to the strains TA98, TA100, WP2 *uvrA* (pKM101), and WP2 (pKM101). This nutrient medium contains per liter:

8 g Nutrient Broth (MERCK, 64293 Darmstadt, Germany)

5 g NaCl (MERCK, 64293 Darmstadt, Germany)

The bacterial cultures were incubated in a shaking water bath for 4 hours at 37° C. The optical density of the bacteria was determined by absorption measurement and the obtained values indicated that the bacteria were harvested at the late exponential or early stationary phase ( $10^8$ - $10^9$  cells/mL).

### 3.3.4 Selective agar

Plates with selective agar (without Histidine/Tryptophan) were used.

### 3.3.5 Overlay agar

The overlay agar contained per litre:

for *Salmonella* strains:

7.0 g Agar Agar\*

6.0 g NaCl\*

10.5 mg L-Histidine×HCl×H<sub>2</sub>O\*

12.2 mg Biotin\*

for *Escherichia coli* strains:

7.0 g Agar Agar\*

6.0 g NaCl\*

10.2 mg Tryptophan\*

\* (MERCK, 64293 Darmstadt, Germany)

Sterilisations were performed at 121° C in an autoclave.

## 3.4 Mammalian Microsomal Fraction S9 Mix

The bacteria used in this assay do not possess the enzyme systems which, in mammals, are known to convert promutagens into active DNA damaging metabolites. In order to overcome this major drawback an exogenous metabolic system is added in the form of mammalian microsome enzyme activation mixture.

### 3.4.1 S9 (Preparation by ICCR-Roßdorf GmbH)

Phenobarbital/β-naphthoflavone induced rat liver S9 was used as the metabolic activation system. The S9 was prepared from male Wistar rats (RjHan:WI; weight approx. 220 – 320 g,



Janvier Labs, 53941 Saint-Berthevin Cedex, France) induced by peroral administration of 80 mg/kg b.w. phenobarbital (Sigma-Aldrich Chemie GmbH, 82024 Taufkirchen, Germany) and by peroral administrations of  $\beta$ -naphthoflavone (Acros Organics, 2440 Geel, Belgium) each, on three consecutive days. The livers were prepared 24 hours after the last treatment. The S9 fractions were produced by dilution of the liver homogenate with a KCl solution (1+3 parts) followed by centrifugation at 9000 g. Aliquots of the supernatant were frozen and stored in ampoules at  $-80^{\circ}\text{C}$ . Small numbers of the ampoules can be kept at  $-20^{\circ}\text{C}$  for up to one week. Each batch of S9 mix is routinely tested with 2-aminoanthracene as well as benzo[a]pyrene (Appendix 3).

The protein concentration in the S9 preparation was 29.6 mg/mL (lot no. 080721D) in both experiments.

### 3.4.2 S9 mix

Before the experiment an appropriate quantity of S9 supernatant was thawed and mixed with S9 cofactor solution. The amount of S9 supernatant was 10% v/v in the S9 mix. Cofactors were added to the S9 mix to reach the following concentrations in the S9 mix:

8 mM  $\text{MgCl}_2$   
33 mM KCl  
5 mM Glucose-6-phosphate  
4 mM NADP

in 100 mM sodium-ortho-phosphate-buffer, pH 7.4.

During the experiment the S9 mix was stored in an ice bath. The S9 mix preparation was performed according to Ames *et al.*(5).

## 3.5 Pre-Experiment for Cytotoxicity

To evaluate the cytotoxicity of the test substance a pre-experiment was performed with all strains. Eight concentrations were tested for cytotoxicity and mutation induction each with three replicate plates. The experimental conditions in this pre-experiment are described in section 3.7 (plate incorporation test).

Cytotoxicity of the test substance results in a reduction in the number of spontaneous revertants (below a factor of 0.5) or a clearing of the bacterial background lawn.

The pre-experiment is reported as the Main Experiment I since the criteria mentioned in Section 3.8.2 Acceptability of the Assay were met.

### 3.6 Concentration Selection

In the pre-experiment the concentration range of the test substance was 3 - 5000 µg/plate. The pre-experiment is reported as Experiment I. Since no cytotoxic effects were observed 5000 µg/plate was chosen as the maximal concentration in Experiment II.

The concentration range included two logarithmic decades. The following concentrations were tested in Experiment II:

33; 100; 333; 1000; 2500; and 5000 µg/plate

### 3.7 Experimental Performance

For each strain and concentration including the controls, three plates were used.

The following materials were mixed in a test tube and poured onto the selective agar plates:

- 100 µL Test solution at each concentration, solvent (negative control) or reference mutagen solution (positive control),
- 500 µL S9 mix (for test with metabolic activation) or S9 mix substitution buffer\* (for test without metabolic activation),
- 100 µL Bacteria suspension (cf. test system, pre-culture of the strains; OD = 1.0 - 1.2; wavelength = 500 nm; approx.  $8 \times 10^8$  cells/mL),
- 2000 µL Overlay agar

For the pre-incubation method test solution (100 µL) (solvent or reference mutagen solution (positive control)), S9 mix / S9 mix substitution buffer\* (500 µL) and bacteria suspension (100 µL) were mixed in a test tube and incubated at  $37^\circ \text{C} \pm 1.5^\circ \text{C}$  for 60 minutes. After pre-incubation overlay agar (2.0 mL,  $45^\circ \text{C}$ ) was added to each tube. The mixture was poured on selective agar plates.

After solidification the plates were incubated upside down for 72 hours at  $37^\circ \text{C} \pm 1.5^\circ \text{C}$  in the dark, plates were then stored at  $4^\circ \text{C}$  until counted (6).

In parallel to each test a sterile control of the test substance was performed and documented in the raw data. Therefore, stock solution (100 µL) and S9 mix / S9 mix substitution buffer\* (500 µL) were mixed with overlay agar (2.0 mL) and poured on minimal agar plates.

\* Substitution buffer: 7 parts of the 100 mM sodium-ortho-phosphate-buffer pH 7.4 with 3 parts of KCl solution 0.15 M

### 3.8 Data Evaluation

#### 3.8.1 Data recording

The colonies were counted using a Petri Viewer with the software program Ames Study Manager (see section 3.9, Major computerized systems). The evaluation unit was connected

to a PC with printer to print out the individual values, the means from the plates for each concentration together with standard deviations and enhancement factors as compared to the spontaneous reversion rates (see tables of results). The print outs are kept with the raw data. Due to precipitation of the test item some test groups were scored manually (as indicated on data tables).

### **3.8.2 Acceptability of the assay**

The *Salmonella typhimurium* and *Escherichia coli* reverse mutation assay is considered acceptable if it meets the following criteria:

- regular background growth in the negative and solvent control
- the spontaneous reversion rates in the negative and solvent control are in the range of the historical data
- the positive control substances should produce an increase above the threshold of twofold (strains TA 98, TA 100, WP2 uvrA (pKM101, and WP2 (pKM101))) or threefold (strains TA 1535 and TA 1537) the revertant colony count of the corresponding solvent control;
- a minimum of five analysable concentrations should be present with at least four showing no signs of toxic effects, evident as a reduction in the number of revertants below the indication factor of 0.5.

### **3.8.3 Evaluation of results**

A test substance is considered as a mutagen if a biologically relevant increase in the number of revertants of twofold or above (strains TA 98, TA 100, WP2 uvrA (pKM101), and WP2 (pKM101)) or of threefold or above (strains TA 1535 and TA 1537) the spontaneous mutation rate of the corresponding solvent control is observed (1).

A concentration dependent increase is considered biologically relevant if the threshold is reached or exceeded at more than one concentration (6).

An increase of revertant colonies equal or above the threshold at only one concentration is judged as biologically relevant if reproduced in an independent second experiment.

A concentration dependent increase in the number of revertant colonies below the threshold is regarded as an indication of a mutagenic potential if reproduced in an independent second experiment. However, whenever the colony counts remain within the historical range of negative and solvent controls, such an increase is not considered biologically relevant.

### **3.8.4 Biometry**

According to the OECD guideline 471, a statistical analysis of the data is not mandatory.

### **3.9 Major Computerized System**

Petri Viewer Sorcerer Colony Counter 3.0 (Instem, Suffolk IP33 3TA, UK) with the software program Ames Study Manager (v1.24) and Ames Archive Manager (v1.01).

## 4.0 RESULTS AND DISCUSSION

The test substance, thiamethoxam SL (A23943A), was assessed for its potential to induce gene mutations in the plate incorporation test (Experiment I) and the pre-incubation test (Experiment II) using *S. typhimurium* strains TA1535, TA1537, TA98, and TA100, and the *E. coli* strains WP2 (pKM101) and WP2 *uvrA* (pKM101).

In the pre-experiment the concentration range of the test substance was 3 - 5000 µg/plate. The pre-experiment is reported as Experiment I. Since no cytotoxic effects were observed 5000 µg/plate was chosen as the maximal concentration in Experiment II. This is the maximum concentration recommended in the OECD test guideline.

The assay was performed with and without liver microsomal activation. Each concentration, including the controls, was tested in triplicate. The concentration range included two logarithmic decades. The test substance was tested at the following concentrations:

Pre-Experiment/Experiment I: 3; 10; 33; 100; 333; 1000; 2500; and 5000 µg/plate

Experiment II: 33; 100; 333; 1000; 2500; and 5000 µg/plate

No precipitation of the test substance occurred in the overlay agar in the test tubes.

Precipitation of the test item in the overlay agar on the incubated agar plates was observed at 5000 µg/plate in Experiment I only.

The plates incubated with the test substance showed normal background growth up to the maximal dose of 5000 µg/plate with and without S9 mix in all strains used.

No cytotoxic effects, evident as a reduction in the number of revertants (below the indication factor of 0.5), occurred in the test groups with and without metabolic activation.

No substantial increase in revertant colony numbers in any of the six tester strains was observed following treatment with thiamethoxam SL (A23943A) at any concentration, neither in the presence nor absence of metabolic activation (S9 mix). There was also no tendency of higher mutation rates with increasing concentrations in the range below the generally acknowledged border of biological relevance.

Appropriate reference mutagens were used as positive controls. They showed a distinct increase in induced revertant colonies.

In experiment II, the data in solvent control of strain WP2 (pKM101) without S9 mix were slightly above our historical control data range. Since this deviation is rather small (300 versus 299 colonies), this effect is considered to be based upon biologically irrelevant fluctuations in the number of colonies.

## 5.0 CONCLUSIONS

In conclusion, it can be stated that during the described mutagenicity tests and under the experimental conditions reported, thiamethoxam SL (A23943A) did not induce gene mutations by base pair changes or frameshifts in the genome of the strains used.

Therefore, thiamethoxam SL (A23943A) is considered to be non-mutagenic in the *Salmonella typhimurium* and *Escherichia coli* reverse mutation assay.

## 6.0 REFERENCES

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## **TABLES SECTION**



**TABLE 1                      Summary of Results Pre-Experiment/Experiment I**

Study Name: 3977311  
 Experiment: 3977311 VV Plate  
 Assay Conditions:

Study Code: ICCR 3977311  
 Date Plated: 04.01.2022  
 Date Counted: 10.01.2022

Metabolic Activation	Test Group	Concentration (per plate)	Revertant Colony Counts (Mean ±SD)					
			TA 1535	TA 1537	TA 98	TA 100	WP2 pKM101	WP2 uvrA pKM101
Without Activation	Deionised water		14 ± 2	16 ± 2	24 ± 6	118 ± 20	298 ± 9	379 ± 20
	Untreated		15 ± 2	12 ± 3	29 ± 7	125 ± 3	337 ± 11	395 ± 5
	Thiamethoxam	3 µg	13 ± 4	12 ± 2	26 ± 9	119 ± 13	305 ± 6	399 ± 4
	SL (A23943A)	10 µg	11 ± 1	13 ± 3	31 ± 8	121 ± 11	319 ± 6	394 ± 6
		33 µg	15 ± 3	10 ± 3	28 ± 8	113 ± 2	304 ± 6	395 ± 5
		100 µg	13 ± 4	10 ± 4	27 ± 6	108 ± 10	313 ± 16	403 ± 12
		333 µg	12 ± 3	14 ± 4	32 ± 8	120 ± 10	318 ± 8	400 ± 4
		1000 µg	9 ± 3	10 ± 4	34 ± 9	135 ± 14	307 ± 2	401 ± 10
		2500 µg	9 ± 2	16 ± 1	29 ± 7	155 ± 14	309 ± 5	406 ± 6
		5000 µg	14 ± 2 <sup>P</sup>	15 ± 3 <sup>P</sup>	26 ± 1 <sup>P</sup>	141 ± 16 <sup>P</sup>	291 ± 35 <sup>P</sup>	322 ± 24 <sup>P</sup>
	NaN3	10 µg	929 ± 276			1509 ± 261		
	4-NOPD	10 µg			505 ± 36			
	4-NOPD	50 µg		85 ± 4				
	MMS	2.0 µL					3154 ± 164	3533 ± 277
With Activation	Deionised water		12 ± 3	15 ± 3	37 ± 10	117 ± 10	312 ± 3	408 ± 17
	Untreated		11 ± 1	17 ± 3	53 ± 8	110 ± 14	343 ± 3	470 ± 7
	Thiamethoxam	3 µg	12 ± 2	16 ± 3	50 ± 4	107 ± 12	328 ± 22	416 ± 18
	SL (A23943A)	10 µg	10 ± 3	14 ± 3	43 ± 7	112 ± 16	325 ± 16	416 ± 5
		33 µg	11 ± 1	15 ± 1	44 ± 12	119 ± 7	312 ± 13	423 ± 8
		100 µg	12 ± 2	15 ± 5	41 ± 9	103 ± 6	314 ± 15	418 ± 7
		333 µg	14 ± 4	18 ± 3	45 ± 9	110 ± 14	320 ± 10	404 ± 4
		1000 µg	19 ± 4	20 ± 4	47 ± 8	148 ± 11	316 ± 11	409 ± 11
		2500 µg	13 ± 1	16 ± 1	45 ± 7	153 ± 17	319 ± 5	398 ± 5
		5000 µg	12 ± 4 <sup>P</sup>	17 ± 2 <sup>P M</sup>	48 ± 11 <sup>P</sup>	150 ± 6 <sup>P</sup>	312 ± 12 <sup>P</sup>	385 ± 16 <sup>P</sup>
	2-AA	2.5 µg	311 ± 31	428 ± 35	2982 ± 326	3785 ± 191		
	2-AA	10.0 µg					940 ± 8	1634 ± 54
Key to Positive Controls			Key to Plate Postfix Codes					
NaN3	sodium azide			P	Precipitate			
2-AA	2-aminoanthracene			M	Manual count			
4-NOPD	4-nitro-o-phenylene-diamine							
MMS	methyl methane sulfonate							

**TABLE 2                      Summary of Results Experiment II**

Study Name: 3977311  
 Experiment: 3977311 HV2 Pre  
 Assay Conditions:

Study Code: ICCR 3977311  
 Date Plated: 18.01.2022  
 Date Counted: 24.01.2022

Metabolic <u>Activation</u>	Test <u>Group</u>	Concen- tration (per plate)	Revertant Colony Counts (Mean ±SD)					
			TA 1535	TA 1537	TA 98	TA 100	WP2 pKM101	WP2 uvrA pKM101
Without Activation	Deionised water		11 ± 4	11 ± 1	29 ± 8	120 ± 12	300 ± 5	371 ± 25
	Untreated		14 ± 3	11 ± 1	30 ± 5	124 ± 8	340 ± 27	344 ± 23
	Thiamethoxam	33 µg	14 ± 3	11 ± 3	27 ± 6	126 ± 14	309 ± 8	344 ± 9
	SL (A23943A)	100 µg	9 ± 1	11 ± 4	32 ± 10	117 ± 7	295 ± 34	400 ± 32
		333 µg	9 ± 2	10 ± 4	26 ± 1	114 ± 2	299 ± 18	364 ± 54
		1000 µg	12 ± 3	17 ± 2	34 ± 7	138 ± 20	310 ± 14	361 ± 12
		2500 µg	15 ± 1	16 ± 5	33 ± 6	127 ± 10	313 ± 9	348 ± 24
		5000 µg	10 ± 3	11 ± 1	39 ± 9	130 ± 8	312 ± 7	319 ± 11
	NaN3	10 µg	1026 ± 74			1763 ± 27		
	4-NOPD	10 µg			579 ± 21			
	4-NOPD	50 µg		120 ± 14				
	MMS	2.0 µL					3055 ± 165	2888 ± 174
With Activation	Deionised water		11 ± 3	15 ± 4	41 ± 4	115 ± 8	311 ± 17	428 ± 25
	Untreated		14 ± 5	16 ± 5	47 ± 10	121 ± 6	340 ± 15	427 ± 19
	Thiamethoxam	33 µg	12 ± 2	17 ± 5	49 ± 12	118 ± 10	320 ± 29	444 ± 24
	SL (A23943A)	100 µg	15 ± 1	17 ± 5	50 ± 10	136 ± 11	321 ± 9	423 ± 23
		333 µg	10 ± 3	15 ± 1	46 ± 9	116 ± 17	340 ± 47	406 ± 22
		1000 µg	15 ± 5	15 ± 6	54 ± 11	151 ± 9	354 ± 48	431 ± 14
		2500 µg	14 ± 4	15 ± 5	55 ± 12	154 ± 9	357 ± 12	392 ± 15
		5000 µg	15 ± 1	16 ± 5	55 ± 7	156 ± 7	319 ± 21	372 ± 5
	2-AA	2.5 µg	261 ± 7	379 ± 29	2164 ± 242	3350 ± 393		
	2-AA	10.0 µg					952 ± 15	1589 ± 72

Key to Positive Controls

NaN3	sodium azide
2-AA	2-aminoanthracene
4-NOPD	4-nitro-o-phenylene-diamine
MMS	methyl methane sulfonate

**TABLE 3                      Pre-Experiment/Experiment I: 3977311 VV Plate Incorporation Without Metabolic Activation**

Study Name: 3977311  
Experiment: 3977311 VV Plate  
Assay Conditions:

Study Code: ICCR 3977311  
Date Plated: 04.01.2022  
Date Counted: 10.01.2022

Without metabolic activation						
Strain	Compound	Concentration per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
TA 1535	Thiamethoxam SL (A23943A)	3 µg	12.7	4.0	0.9	9, 17, 12
		10 µg	11.0	1.0	0.8	10, 11, 12
		33 µg	14.7	2.5	1.1	17, 15, 12
		100 µg	12.7	4.0	0.9	12, 17, 9
		333 µg	12.0	2.6	0.9	15, 11, 10
		1000 µg	8.7	2.9	0.6	7, 7, 12
		2500 µg	9.0	1.7	0.7	10, 10, 7
		5000 µg	14.0	2.0	1.0	12 P, 16 P, 14 P
	Deionised water		13.7	1.5		12, 14, 15
	Untreated		15.3	1.5		17, 14, 15
TA 1537	Thiamethoxam SL (A23943A)	3 µg	12.0	1.7	0.8	11, 14, 11
		10 µg	13.3	2.9	0.9	15, 15, 10
		33 µg	10.3	2.9	0.7	7, 12, 12
		100 µg	10.3	3.5	0.7	7, 14, 10
		333 µg	14.0	3.6	0.9	10, 17, 15
		1000 µg	10.3	3.5	0.7	7, 10, 14
		2500 µg	16.3	0.6	1.0	17, 16, 16
		5000 µg	14.7	3.2	0.9	16 P, 11 P, 17 P
	Deionised water		15.7	1.5		14, 17, 16
	Untreated		12.0	3.0		12, 9, 15
TA 98	Thiamethoxam SL (A23943A)	3 µg	25.7	9.0	1.1	36, 20, 21
		10 µg	31.0	7.8	1.3	40, 27, 26
		33 µg	28.3	8.1	1.2	27, 21, 37
		100 µg	26.7	5.7	1.1	25, 22, 33
		333 µg	31.7	8.1	1.3	26, 41, 28
		1000 µg	34.3	9.1	1.4	44, 26, 33
		2500 µg	29.0	6.6	1.2	22, 35, 30
		5000 µg	26.0	1.0	1.1	27 P, 25 P, 26 P
	Deionised water		24.3	6.1		23, 19, 31
	Untreated		29.3	7.1		37, 23, 28
Key to Plate Postfix Codes						
P      Precipitate						

Study Name: 3977311  
 Experiment: 3977311 VV Plate  
 Assay Conditions:

Study Code: ICCR 3977311  
 Date Plated: 04.01.2022  
 Date Counted: 10.01.2022

**Without metabolic activation**

Strain	Compound	Concentration per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
<b>TA 100</b>	<b>Thiamethoxam SL (A23943A)</b>	3 µg	119.3	12.5	1.0	107, 132, 119
		10 µg	121.3	10.6	1.0	123, 131, 110
		33 µg	112.7	2.1	1.0	115, 112, 111
		100 µg	108.3	10.4	0.9	120, 105, 100
		333 µg	120.3	10.4	1.0	132, 112, 117
		1000 µg	135.3	13.5	1.2	135, 122, 149
		2500 µg	155.3	13.6	1.3	141, 157, 168
		5000 µg	140.7	15.5	1.2	136 P, 128 P, 158 P
	<b>Deionised water</b>		117.7	19.9		122, 96, 135
	<b>Untreated</b>		125.0	3.0		125, 128, 122
<b>WP2 pKM101</b>	<b>Thiamethoxam SL (A23943A)</b>	3 µg	305.3	6.1	1.0	312, 300, 304
		10 µg	318.7	6.4	1.1	326, 315, 315
		33 µg	303.7	5.5	1.0	310, 300, 301
		100 µg	313.3	16.1	1.1	325, 320, 295
		333 µg	318.0	8.2	1.1	327, 316, 311
		1000 µg	306.7	2.1	1.0	306, 305, 309
		2500 µg	309.3	4.9	1.0	306, 315, 307
		5000 µg	291.3	34.9	1.0	311 P, 251 P, 312 P
	<b>Deionised water</b>		298.3	8.5		298, 307, 290
	<b>Untreated</b>		336.7	10.7		349, 330, 331
<b>WP2 uvrA pKM101</b>	<b>Thiamethoxam SL (A23943A)</b>	3 µg	399.3	3.5	1.1	396, 399, 403
		10 µg	394.0	6.0	1.0	394, 400, 388
		33 µg	394.7	4.7	1.0	391, 393, 400
		100 µg	403.3	12.3	1.1	417, 400, 393
		333 µg	399.7	4.0	1.1	399, 404, 396
		1000 µg	401.3	10.3	1.1	390, 410, 404
		2500 µg	406.3	5.5	1.1	400, 409, 410
		5000 µg	321.7	24.0	0.8	336 P, 335 P, 294 P
	<b>Deionised water</b>		379.3	20.1		385, 396, 357
	<b>Untreated</b>		394.7	4.5		390, 395, 399
<b>TA 1535</b>	<b>NaN3</b>	10 µg	929.0	276.3	68.0	1198, 646, 943
<b>TA 1537</b>	<b>4-NOPD</b>	50 µg	85.3	4.2	5.4	90, 82, 84
<b>TA 98</b>	<b>4-NOPD</b>	10 µg	505.0	35.8	20.8	521, 530, 464
<b>TA 100</b>	<b>NaN3</b>	10 µg	1509.0	261.1	12.8	1245, 1515, 1767
<b>WP2 pKM101</b>	<b>MMS</b>	2.0 µL	3154.0	163.8	10.6	3104, 3337, 3021
<b>WP2 uvrA pKM101</b>	<b>MMS</b>	2.0 µL	3532.7	277.3	9.3	3393, 3353, 3852
Key to Positive Controls						Key to Plate Postfix Codes
NaN3	sodium azide					P Precipitate
4-NOPD	4-nitro-o-phenylene-diamine					
MMS	methyl methane sulfonate					

**TABLE 4                      Pre-Experiment/Experiment I: 3977311 VV Plate Incorporation  
With Metabolic Activation**

Study Name: 3977311  
Experiment: 3977311 VV Plate  
Assay Conditions:

Study Code: ICCR 3977311  
Date Plated: 04.01.2022  
Date Counted: 10.01.2022

**With metabolic activation**

Strain	Compound	Concen- tration per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
TA 1535	Thiamethoxam SL (A23943A)	3 µg	12.3	1.5	1.1	14, 12, 11
		10 µg	10.3	2.9	0.9	12, 7, 12
		33 µg	11.0	1.0	0.9	12, 10, 11
		100 µg	12.0	1.7	1.0	14, 11, 11
		333 µg	14.3	3.8	1.2	16, 10, 17
		1000 µg	19.0	3.6	1.6	20, 22, 15
		2500 µg	13.3	1.2	1.1	12, 14, 14
		5000 µg	12.0	3.6	1.0	11 P, 16 P, 9 P
	Deionised water		11.7	3.1		9, 15, 11
	Untreated		10.7	0.6		10, 11, 11
TA 1537	Thiamethoxam SL (A23943A)	3 µg	16.0	3.5	1.1	14, 20, 14
		10 µg	14.0	2.6	1.0	11, 15, 16
		33 µg	15.0	1.0	1.0	14, 15, 16
		100 µg	15.3	5.0	1.0	10, 16, 20
		333 µg	18.3	3.1	1.2	15, 21, 19
		1000 µg	20.3	4.2	1.4	25, 17, 19
		2500 µg	16.3	0.6	1.1	17, 16, 16
		5000 µg	17.3	1.5	1.2	16 P M, 19 P M, 17 P M
	Deionised water		14.7	2.5		12, 15, 17
	Untreated		17.3	2.5		17, 15, 20
TA 98	Thiamethoxam SL (A23943A)	3 µg	50.0	3.6	1.4	54, 49, 47
		10 µg	42.7	6.7	1.2	35, 46, 47
		33 µg	44.0	12.2	1.2	36, 58, 38
		100 µg	40.7	9.3	1.1	38, 51, 33
		333 µg	45.0	8.7	1.2	49, 51, 35
		1000 µg	47.3	7.6	1.3	42, 44, 56
		2500 µg	45.0	7.0	1.2	42, 40, 53
		5000 µg	47.7	10.5	1.3	37 P, 48 P, 58 P
	Deionised water		37.0	10.4		43, 25, 43
	Untreated		53.0	7.5		61, 52, 46

Key to Plate Postfix Codes

P      Precipitate  
M      Manual count

Study Name: 3977311  
 Experiment: 3977311 VV Plate  
 Assay Conditions:

Study Code: ICCR 3977311  
 Date Plated: 04.01.2022  
 Date Counted: 10.01.2022

**With metabolic activation**

Strain	Compound	Concentration per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
<b>TA 100</b>	<b>Thiamethoxam SL (A23943A)</b>	3 µg	107.3	11.9	0.9	102, 121, 99
		10 µg	112.3	15.6	1.0	127, 96, 114
		33 µg	118.7	6.7	1.0	122, 111, 123
		100 µg	103.3	6.4	0.9	96, 107, 107
		333 µg	109.7	13.9	0.9	106, 98, 125
		1000 µg	147.7	10.5	1.3	137, 148, 158
		2500 µg	153.0	17.3	1.3	172, 138, 149
		5000 µg	149.7	5.9	1.3	152 P, 143 P, 154 P
	<b>Deionised water</b>		116.7	10.4		105, 125, 120
	<b>Untreated</b>		110.0	14.1		112, 95, 123
<b>WP2 pKM101</b>	<b>Thiamethoxam SL (A23943A)</b>	3 µg	328.0	22.3	1.1	353, 321, 310
		10 µg	325.3	15.6	1.0	340, 327, 309
		33 µg	312.3	12.7	1.0	305, 305, 327
		100 µg	314.0	14.8	1.0	331, 307, 304
		333 µg	320.3	10.0	1.0	321, 330, 310
		1000 µg	315.7	10.5	1.0	326, 316, 305
		2500 µg	318.7	4.7	1.0	324, 317, 315
		5000 µg	312.0	12.3	1.0	317 P, 298 P, 321 P
	<b>Deionised water</b>		312.0	3.0		315, 309, 312
	<b>Untreated</b>		343.0	2.6		341, 342, 346
<b>WP2 uvrA pKM101</b>	<b>Thiamethoxam SL (A23943A)</b>	3 µg	416.0	18.4	1.0	429, 395, 424
		10 µg	416.0	5.0	1.0	421, 416, 411
		33 µg	423.0	8.2	1.0	416, 421, 432
		100 µg	418.0	7.2	1.0	426, 412, 416
		333 µg	403.7	4.0	1.0	406, 399, 406
		1000 µg	409.0	10.5	1.0	420, 408, 399
		2500 µg	398.3	4.6	1.0	401, 393, 401
		5000 µg	385.3	16.2	0.9	368 P, 388 P, 400 P
	<b>Deionised water</b>		408.3	17.0		389, 415, 421
	<b>Untreated</b>		469.7	6.7		477, 468, 464
<b>TA 1535</b>	<b>2-AA</b>	2.5 µg	310.7	31.4	26.6	300, 346, 286
<b>TA 1537</b>	<b>2-AA</b>	2.5 µg	428.3	35.2	29.2	391, 461, 433
<b>TA 98</b>	<b>2-AA</b>	2.5 µg	2982.0	326.3	80.6	2607, 3138, 3201
<b>TA 100</b>	<b>2-AA</b>	2.5 µg	3785.3	190.6	32.4	3949, 3831, 3576
<b>WP2 pKM101</b>	<b>2-AA</b>	10.0 µg	940.3	8.3	3.0	931, 947, 943
<b>WP2 uvrA pKM101</b>	<b>2-AA</b>	10.0 µg	1634.3	54.2	4.0	1650, 1574, 1679

Key to Positive Controls

2-AA 2-aminoanthracene

Key to Plate Postfix Codes

P Precipitate  
 M Manual count

**TABLE 5 Experiment II: 3977311 HV2 Pre Incubation Without Metabolic Activation**

Study Name: 3977311  
Experiment: 3977311 HV2 Pre  
Assay Conditions:

Study Code: ICCR 3977311  
Date Plated: 18.01.2022  
Date Counted: 24.01.2022

Without metabolic activation						
Strain	Compound	Concentration per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
TA 1535	Thiamethoxam SL (A23943A)	33 µg	13.7	2.5	1.3	14, 16, 11
		100 µg	9.3	0.6	0.9	9, 9, 10
		333 µg	9.0	2.0	0.8	11, 7, 9
		1000 µg	11.7	2.9	1.1	10, 10, 15
		2500 µg	15.0	1.0	1.4	15, 16, 14
		5000 µg	10.0	2.6	0.9	7, 12, 11
	Deionised water		10.7	4.0		15, 10, 7
	Untreated		13.7	2.5		11, 14, 16
TA 1537	Thiamethoxam SL (A23943A)	33 µg	11.3	3.2	1.0	10, 9, 15
		100 µg	10.7	3.5	0.9	11, 14, 7
		333 µg	9.7	4.0	0.9	14, 9, 6
		1000 µg	17.0	2.0	1.5	15, 17, 19
		2500 µg	16.0	4.6	1.4	17, 20, 11
		5000 µg	10.7	1.2	0.9	12, 10, 10
	Deionised water		11.3	1.2		12, 12, 10
	Untreated		10.7	0.6		10, 11, 11
TA 98	Thiamethoxam SL (A23943A)	33 µg	26.7	5.7	0.9	33, 25, 22
		100 µg	32.3	10.0	1.1	42, 33, 22
		333 µg	26.3	1.2	0.9	27, 25, 27
		1000 µg	34.0	7.2	1.2	32, 28, 42
		2500 µg	33.3	5.5	1.1	27, 37, 36
		5000 µg	38.7	8.6	1.3	48, 37, 31
	Deionised water		29.3	8.1		38, 28, 22
	Untreated		30.3	5.0		35, 25, 31
TA 100	Thiamethoxam SL (A23943A)	33 µg	126.0	14.0	1.1	136, 132, 110
		100 µg	117.0	7.0	1.0	122, 120, 109
		333 µg	114.0	2.0	1.0	114, 116, 112
		1000 µg	138.0	19.7	1.2	154, 116, 144
		2500 µg	127.0	9.5	1.1	116, 132, 133
		5000 µg	129.7	7.6	1.1	133, 135, 121
	Deionised water		120.0	12.2		128, 126, 106
	Untreated		124.0	8.2		122, 117, 133
WP2 pKM101	Thiamethoxam SL (A23943A)	33 µg	309.0	8.2	1.0	316, 311, 300
		100 µg	295.0	33.8	1.0	326, 300, 259
		333 µg	299.3	17.5	1.0	299, 282, 317
		1000 µg	310.0	14.2	1.0	299, 305, 326
		2500 µg	313.0	9.2	1.0	321, 303, 315
		5000 µg	312.3	7.1	1.0	320, 311, 306
	Deionised water		299.7	4.7		305, 298, 296
	Untreated		340.3	26.5		370, 332, 319

Study Name: 3977311  
 Experiment: 3977311 HV2 Pre  
 Assay Conditions:

Study Code: ICCR 3977311  
 Date Plated: 18.01.2022  
 Date Counted: 24.01.2022

**Without metabolic activation**

Strain	Compound	Concentration per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
<b>WP2 uvrA pKM101</b>	<b>Thiamethoxam SL (A23943A)</b>	33 µg	343.7	9.0	0.9	353, 343, 335
		100 µg	399.7	32.3	1.1	374, 389, 436
		333 µg	364.3	53.8	1.0	426, 340, 327
		1000 µg	361.3	12.2	1.0	372, 364, 348
		2500 µg	347.7	23.9	0.9	375, 337, 331
		5000 µg	318.7	11.0	0.9	325, 306, 325
	<b>Deionised water</b>		371.3	24.8		351, 364, 399
	<b>Untreated</b>		343.7	23.4		317, 353, 361
<b>TA 1535</b>	<b>NaN3</b>	10 µg	1025.7	73.7	96.2	1093, 1037, 947
<b>TA 1537</b>	<b>4-NOPD</b>	50 µg	120.0	14.2	10.6	131, 125, 104
<b>TA 98</b>	<b>4-NOPD</b>	10 µg	578.7	20.6	19.7	600, 577, 559
<b>TA 100</b>	<b>NaN3</b>	10 µg	1763.3	26.7	14.7	1788, 1735, 1767
<b>WP2 pKM101</b>	<b>MMS</b>	2.0 µL	3055.0	165.0	10.2	2870, 3108, 3187
<b>WP2 uvrA pKM101</b>	<b>MMS</b>	2.0 µL	2888.0	173.6	7.8	2934, 2696, 3034

Key to Positive Controls

NaN3	sodium azide
4-NOPD	4-nitro-o-phenylene-diamine
MMS	methyl methane sulfonate



**TABLE 6 Experiment II: 3977311 HV2 Pre Incubation With Metabolic Activation**

Study Name: 3977311  
Experiment: 3977311 HV2 Pre  
Assay Conditions:

Study Code: ICCR 3977311  
Date Plated: 18.01.2022  
Date Counted: 24.01.2022

With metabolic activation						
Strain	Compound	Concentration per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
TA 1535	Thiamethoxam SL (A23943A)	33 µg	12.0	1.7	1.1	11, 11, 14
		100 µg	15.0	1.0	1.3	15, 14, 16
		333 µg	9.7	3.2	0.9	12, 6, 11
		1000 µg	14.7	4.6	1.3	12, 20, 12
		2500 µg	14.0	3.6	1.2	10, 17, 15
		5000 µg	14.7	0.6	1.3	14, 15, 15
	Deionised water		11.3	3.2		15, 10, 9
	Untreated		14.3	4.6		17, 9, 17
TA 1537	Thiamethoxam SL (A23943A)	33 µg	17.3	4.9	1.2	23, 15, 14
		100 µg	17.0	5.3	1.1	11, 21, 19
		333 µg	15.0	1.0	1.0	14, 16, 15
		1000 µg	15.0	5.6	1.0	9, 16, 20
		2500 µg	15.3	4.5	1.0	20, 11, 15
		5000 µg	16.3	4.5	1.1	21, 12, 16
	Deionised water		15.0	3.6		14, 12, 19
	Untreated		16.3	4.5		16, 12, 21
TA 98	Thiamethoxam SL (A23943A)	33 µg	48.7	11.9	1.2	57, 35, 54
		100 µg	50.3	9.6	1.2	59, 40, 52
		333 µg	45.7	8.5	1.1	54, 37, 46
		1000 µg	54.0	11.3	1.3	67, 47, 48
		2500 µg	54.7	12.2	1.3	52, 44, 68
		5000 µg	54.7	7.1	1.3	47, 56, 61
	Deionised water		40.7	3.5		44, 41, 37
	Untreated		47.0	10.1		56, 36, 49
TA 100	Thiamethoxam SL (A23943A)	33 µg	118.3	10.3	1.0	107, 127, 121
		100 µg	136.0	10.6	1.2	148, 128, 132
		333 µg	116.0	17.3	1.0	107, 136, 105
		1000 µg	151.0	8.9	1.3	144, 161, 148
		2500 µg	154.0	8.9	1.3	161, 157, 144
		5000 µg	156.3	7.0	1.4	163, 149, 157
	Deionised water		114.7	7.8		117, 106, 121
	Untreated		121.3	6.4		125, 114, 125
WP2 pKM101	Thiamethoxam SL (A23943A)	33 µg	320.3	29.3	1.0	306, 301, 354
		100 µg	321.0	8.7	1.0	325, 311, 327
		333 µg	339.7	47.2	1.1	295, 335, 389
		1000 µg	353.7	47.9	1.1	406, 312, 343
		2500 µg	357.0	12.5	1.1	343, 367, 361
		5000 µg	319.0	20.8	1.0	331, 295, 331
	Deionised water		311.3	16.8		326, 315, 293
	Untreated		340.0	14.5		354, 341, 325

Study Name: 3977311  
 Experiment: 3977311 HV2 Pre  
 Assay Conditions:

Study Code: ICCR 3977311  
 Date Plated: 18.01.2022  
 Date Counted: 24.01.2022

**With metabolic activation**

Strain	Compound	Concentration per plate	Mean revertants per plate	Standard Deviation	Ratio treated / solvent	Individual revertant colony counts
<b>WP2 uvrA pKM101</b>	<b>Thiamethoxam SL (A23943A)</b>	33 µg	443.7	23.7	1.0	469, 422, 440
		100 µg	423.0	22.6	1.0	442, 429, 398
		333 µg	405.7	21.5	0.9	430, 389, 398
		1000 µg	431.0	14.0	1.0	431, 445, 417
		2500 µg	392.3	14.6	0.9	394, 406, 377
		5000 µg	371.7	4.7	0.9	377, 368, 370
	<b>Deionised water</b>		428.3	24.5		452, 403, 430
	<b>Untreated</b>		427.0	19.0		427, 446, 408
<b>TA 1535</b>	<b>2-AA</b>	2.5 µg	261.3	6.7	23.1	258, 269, 257
<b>TA 1537</b>	<b>2-AA</b>	2.5 µg	378.7	29.3	25.2	352, 374, 410
<b>TA 98</b>	<b>2-AA</b>	2.5 µg	2163.7	242.0	53.2	1910, 2392, 2189
<b>TA 100</b>	<b>2-AA</b>	2.5 µg	3349.7	392.9	29.2	3576, 3577, 2896
<b>WP2 pKM101</b>	<b>2-AA</b>	10.0 µg	951.7	15.0	3.1	935, 956, 964
<b>WP2 uvrA pKM101</b>	<b>2-AA</b>	10.0 µg	1588.7	71.6	3.7	1515, 1658, 1593

Key to Positive Controls

2-AA      2-aminoanthracene

## **APPENDICES SECTION**

## APPENDIX 1 Historical Control Data

These data represent the laboratory's historical control data from July 2018 until July 2020 representing approx. 600 experiments (WP2 pKM101, WP2 uvrA pKM101 the historical data are based on approx. 80 experiments).

The plate incorporation and preincubation methods were used to establish the historical control data since the bacterial strains used in the Ames test do not show differences in their revertant numbers beyond the range of the biological variability in the controls due to the different assay types

The positive controls that used to compile the historical positive control data correspond to the positive control substances described in Methods; section 3.2.2 (Positive control substances).

Strain		without S9 mix				with S9 mix			
		Mean	SD	Min	Max	Mean	SD	Min	Max
TA 1535	Solvent control	12	2.6	7	22	13	2.5	7	24
	Untreated control	12	2.9	6	26	13	2.8	7	23
	Positive control	1116	141.3	340	1612	346	72.1	170	736
TA1537	Solvent control	11	2.4	6	20	14	2.8	7	28
	Untreated control	11	2.8	5	22	14	3.2	7	30
	Positive control	83	22.1	48	400	286	98.7	82	630
TA 98	Solvent control	28	4.9	13	46	38	6.4	12	62
	Untreated control	29	5.0	14	48	41	6.8	14	64
	Positive control	421	91.2	216	1218	3275	774.9	322	5699
TA 100	Solvent control	127	30.7	63	214	131	30.0	72	214
	Untreated control	135	35.7	64	233	140	34.4	68	217
	Positive control	1759	273.4	511	2588	3566	837.6	553	5444
WP2 pKM 101	Solvent control	248	31.7	171	299	266	33.0	205	315
	Untreated control	269	26.6	212	346	299	28.2	233	345
	Positive control	3343	428.4	2332	4653	1092	257.8	933	2781
WP2uvrA pKM 101	Solvent control	322	31.6	248	388	375	38.5	287	466
	Untreated control	346	28.2	279	403	393	32.6	313	480
	Positive control	3176	468.5	2021	4717	1897	183.2	1270	2464

Mean = mean value of revertants/plate

SD = standard deviation

Min = minimal value

Max = maximal value

## APPENDIX 2 Copy of GLP Certificate



### Gute Laborpraxis/Good Laboratory Practice

### GLP-Bescheinigung/Statement of GLP Compliance

(gemäß/according to § 19b Abs. 1 Chemikaliengesetz)

HESSEN



Eine GLP-Inspektion zur Überwachung der Einhaltung der GLP-Grundsätze gemäß Chemikaliengesetz bzw. Richtlinie 2004/9/EG wurde durchgeführt in

Assessment of conformity with GLP according to Chemikaliengesetz and Directive 2004/9/EEC at:

☒ Prüfeinrichtung/Test facility

☐ Prüfstandort/Test site

#### ICCR-Roßdorf GmbH

Institute for Competent Contract Research  
In den Leppsteinswiesen 19  
64380 Roßdorf

(Unverwechselbare Bezeichnung und Adresse/Unequivocal name and address)

#### Prüfungen nach Kategorien/Areas of Expertise

(gemäß/according to ChemVwV-GLP Nr. 5.3/OECD guidance)

2 Prüfungen zur Bestimmung der toxikologischen Eigenschaften  
3 Prüfungen zur Bestimmung der erbgutverändernden Eigenschaften (in vitro und in vivo)  
8 Analytische Prüfungen an biologischen Materialien

2 Toxicity studies  
3 Mutagenicity studies  
8 Analytical and clinical chemistry testing

22.11.2018, 21.02.2019, 12. bis 14.03.2019

Datum der Inspektion/Date of Inspection  
(Tag Monat Jahr/day month year)

Die genannte Prüfeinrichtung befindet sich im nationalen GLP-Überwachungsverfahren und wird regelmäßig auf Einhaltung der GLP-Grundsätze überwacht.

The above mentioned test facility is included in the national GLP Compliance Programme and is inspected on a regular basis.

Auf der Grundlage des Inspektionsberichtes wird hiermit bestätigt, dass in dieser Prüfeinrichtung die oben genannten Prüfungen unter Einhaltung der GLP-Grundsätze durchgeführt werden können.

Based on the inspection report it can be confirmed, that this test facility is able to conduct the aforementioned studies in compliance with the Principles of GLP.

Im Auftrag

Dr. Astrid Brandt, Referentin, Wiesbaden, den 23. Oktober 2019  
(Name und Funktion der verantwortlichen Person/  
Name and function of responsible person)



Hessisches Ministerium für Umwelt, Klimaschutz, Landwirtschaft und Verbraucherschutz,  
Mainzer Straße 80, D 65189 Wiesbaden

(Name und Adresse der GLP-Überwachungsbehörde/Name and address of the GLP Monitoring Authority)

English name and address of the GLP Monitoring Authority: Hessian Ministry for Environment, Climate Protection, Agriculture and Consumer Protection; Department II 10; P.O. Box 31 09; 65189 Wiesbaden

Translation of seal inscription: Hessian Ministry for Environment, Climate Protection, Agriculture and Consumer Protection

## APPENDIX 3 Certificate of S9



### CERTIFICATE

ICCR-Roßdorf S9 Preparation Lot No. 080721D

Date of preparation: July 08, 2021

Release date: July 26, 2021

Protein assay: 29.6 mg protein / ml S9

Sterility: 7.6 colonies / ml S9 on glucose-minimal-agar

Salmonella typhimurium assay (AMES-test)

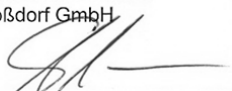
Treatment	µl S9 / plate	number of revertants in TA 98
negative	0	33
control	100	54
10 µg/plate	0	40
2-Aminoanthracene	100	2031
10 µg/plate	0	23
Benzo(a)pyrene	100	88

The S9 was obtained from the livers of male Wistar rats which received triple treatments of 80 mg / kg body weight Phenobarbital and  $\beta$ -Naphthoflavone orally on consecutive days. The livers were prepared 24 hours after the last treatment.

  
Quality Assurance Auditor  
ICCR-Roßdorf GmbH

27. JULI 2021

Date

  
Dr. Steffen Naumann  
Study Director  
ICCR-Roßdorf GmbH

27. JULI 2021

Date

ICCR-Roßdorf GmbH  
In den Leppsteinswiesen 19, 64380 Roßdorf, Deutschland  
T +49 6154 8070 F +49 6154 83399  
Registriergericht Darmstadt, HRB 6837, USt-ID DE812333696  
Geschäftsführer: Dr. Markus Schulz

SOP Origin TS-SOP S9\_23

Best available copy

## APPENDIX 4 Certificate of Analysis



ALS Laboratórios LS Ltda.  
Rua Fábria, 59 – CEP: 05051-030  
São Paulo, SP - Brazil

SYNGENTA PROTEÇÃO DE CULTIVOS Ltda.  
Rua Doutor Rubens Gomes Bueno nº 691,  
11º andar, Torre Sigma  
CEP 04730-000 – Bairro Várzea de Baixo  
São Paulo-SP – Brazil

### Certificate of Analysis

**A23943A**  
**Thiamethoxam SL (075)**  
**NSI001-085-017**

<b>Batch Identification</b>	NSI001-085-017
<b>Product Code</b>	A23943A
<b>Other Product Code(s)</b>	A23943A; EXF23867A; CGA293343 SL (075); Thiamethoxam SL (075)
<b>EUP number:</b>	739/2021 Expiry date: 26/04/2024
<b>Received on:</b>	28 September 2021
<b>Source</b>	Syngenta Proteção de Cultivos Ltda. Rodovia Professor Zeferino Vaz, SP 332, s/nº, km 127,5 – Bairro Santa Terezinha, CEP 13148-915 – Paulínia-SP – Brazil

#### Chemical Analysis (Active Ingredients Content)

– **Content of Thiamethoxam \*** **6.63 % w/w corresponding to 75.42 g/L**

The Active Ingredient content is within the FAO limits.  
Methodology used for Characterization: HPLC (SF-1151/1)

#### Physical Analysis

– <b>Appearance</b>	Homogeneous and translucent
– <b>Color</b>	6/12 – 5YR (Dark Orange)
– <b>Physical state</b>	Liquid
– <b>Density *</b>	1.1378 g/cm <sup>3</sup>

#### Stability:

– <b>Storage Temperature</b>	<30°C
– <b>Recertification Date</b>	10 September 2023

If stored under the conditions given above, this test item can be considered stable until the recertification date is reached.

This Certificate of Analysis summarizes data which originates either from a single study or from several individual studies. Tests marked with an asterisk (\*) have been conducted in compliance with GLP. All original raw data, including any storage medium for electronically recorded data, documentation, the signed study plan, the protocol amendments, the final report and a sample of the test item will be retained in the GLP Archives at ALS Laboratórios LS Ltda.

Study number of batch characterization: 26785/2021CF and 26787/2021CC

Authorization: 10 November 2021

*Victor F. G. da Silva*  
Victor Ferreira Gomes da Silva  
ALS Laboratórios LS Ltda.

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